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Fecal microbiota transplantation combined with ruxolitinib as a salvage treatment for intestinal steroid-refractory acute GVHD

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Abstract

Acute graft-versus-host disease (aGVHD), especially intestinal aGVHD, is one of the most severe complications after allogeneic hematopoietic stem cell transplantation (HSCT). Fecal microbiota transplantation (FMT) has been applied to the treatment of intestinal steroid-refractory aGVHD (SR-aGVHD). Ruxolitinib is the first drug recommended for SR-aGVHD. Here, we reported the outcome data from 21 patients who had received the combined treatment of FMT with ruxolitinib as a salvage treatment in intestinal SR-aGVHD after HSCT. The overall response rate on day 28 was 71.4% (95% CI 50.4–92.5%), including 10 patients with complete responses. The durable overall response at day 56 in responders was 80%. GVHD relapse rate was 33.3% in responders. The levels of inflammatory cytokines as well as T cells and NK cells activation declined. The diversity of the intestinal microbiota was improved in responders. Viral reactivations and severe cytopenia were the major adverse events (61.9% and 81% respectively). The estimated 6-month overall survival was 57.1% (95% CI: 35.9–78.3%), while event-free survival was 52.4% (95% CI: 21.7%–64.1%). Collectively, FMT with ruxolitinib could be an effective treatment for intestinal SR-aGVHD after HSCT.

Trial registration: ClinicalTrials.gov identifier: NCT03148743.

Keywords: HSCT, FMT, Ruxolitinib, Steroid-refractory GVHD, Intestinal GVHD

To the Editor,

Acute graft-versus-host disease (aGVHD), especially intestinal localization, remains one of the most unremovable barriers to the success of allogeneic hematopoietic stem cell transplantation (HSCT), leading to late morbidity and mortality. Fecal microbiota transplantation (FMT) was reported to be effective [1], but attempts have been made to explore combination treatments with other

drugs to increase response rate and improve survival of intestinal steroid-refractory aGVHD (SR-aGVHD). The initial experience reported by Bilinski et al. [2], as well as the clinical trial (*ClinicalTrials.gov ID: NCT04269850*) sponsored by St. Petersburg State Pavlov Medical University [3], gave some evidence to show the potential efficiency of the combined treatment with FMT and ruxolitinib. We previously conducted a phase 1 clinical trial of FMT as a therapeutic option for intestinal GVHD (*ClinicalTrials.gov ID: NCT03148743*) [4–6] and mentioned a subset of patients using ruxolitinib with a higher response rate. Herein, we reported this subset showing the efficacy of combined treatment of FMT with ruxolitinib as a salvage treatment in intestinal SR-aGVHD after HSCT.

A cohort of 21 patients was treated with FMT and the add-on ruxolitinib as an additional therapy for grade

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Table 1 Patient and treatment characteristics

#	Age (years)	Gender	Diagnose	Disease status	Donor source	Conditioning regimen	Stem cell source	GVHD Prophylaxis	Onset of aGVHD after HSCT
1	44	Male	AML	CR	MUD(O → AB)	Bu/Cy	PB	CsA + MMF + MTX + ATG	22
2	59	Male	AML	Relapse	HRD(A → A)	CBA	BM + PB	CsA + MMF + MTX + ATG	19
3	34	Male	AML	CR	HRD(O → B)	Bu/Cy	BM + PB	CsA + MMF + MTX + ATG	19
4	29	Female	AML	Refractory	HRD(B → B)	Bu/Cy	BM + PB	CsA + MMF + MTX + ATG	10
5	40	Male	AML	Relapse	HRD(B → A)	Bu/Cy	BM + PB	CsA + MMF + MTX + ATG	30
6	18	Male	SAA	NR	HRD(A → B)	Bu/Cy	BM + PB	CsA + MMF + MTX + ATG	20
7	29	Male	AML	CR	HRD(B → B)	Bu/Cy	BM + PB	CsA + MMF + MTX + ATG	30
8	18	Female	ALL	CR	HRD(B → B)	Bu/Cy	BM + PB	CsA + MMF + MTX + ATG	25
9	47	Male	AML	CR	HRD(O → A)	Bu/Cy	BM + PB	CsA + MMF + MTX + ATG	97
10	21	Female	ALL	CR	HRD(O → B)	Bu/Cy	BM + PB	CsA + MMF + MTX + ATG	42
11	28	Female	AML	CR	HRD(A → A)	Bu/Cy	BM + PB	FK506 + MMF + MTX + ATG	22
12	47	Male	MDS	PD	HRD(B → O)	Bu/Cy	BM + PB	CsA + MMF + MTX + ATG	17
13	16	Male	SAA	NR	HRD(AB → AB)	Bu/Cy	BM + PB	CsA + MMF + MTX + ATG	18
14	23	Female	AML	CR	HRD(A → A)	Bu/Cy	BM + PB	CsA + MMF + MTX + ATG	22
15	38	Male	AML	CR	HRD(B → B)	Bu/Cy	PB	CsA + MMF + MTX + ATG	55
16	15	Female	MDS	NR	HRD(B → B)	Bu/Cy	BM + PB	CsA + MMF + MTX + ATG	24
17	25	Male	AML	Relapse	HRD(O → B)	Bu/Cy	BM + PB	CsA + MMF + MTX + ATG	51
18	45	Male	HAL	CR	HRD(O → O)	Bu/Cy	BM + PB	CsA + MMF + MTX + ATG	100
19	16	Male	ALL	Refractory	HRD(A → A)	Bu/Cy	BM + PB	CsA + MMF + MTX + ATG	28
20	26	Male	MDS	PD	HRD(AB → AB)	Bu/Cy	BM + PB	CsA + MMF + MTX + ATG	23
21	54	Male	AML	CR	HRD(O → O)	Bu/Cy	BM + PB	CsA + MMF + MTX + ATG	22

#	Additional organ involvement to intestine manifestations	aGVHD grade	Response to combined treatment	Virus reactivation after combined treatment	Infection after combined treatment	Severe cytopenia	Last follow-up (days after response)	Status	GVHD relapse (severity, days after response)
1	None	III	NR	CMV	–	3	111	Dead	–
2	Skin and liver	IV	NR	–	–	4	40	Dead	–
3	Skin	IV	PR	–	–	4	127	Dead	Intestine (1, +47d)
4	Skin and liver	IV	NR	–	–	4	22	Dead	–
5	Skin and liver	III	PR	CMV	–	–	73	Dead	–
6	None	III	CR	CMV enteritis	–	–	677	Alive	–
7	Skin and liver	III	PR	–	–	4	661	Alive	Skin, intestine (2, +88d)
8	Skin	III	PR	CMV enteritis, EBV	–	–	685	Alive	–
9	None	III	CR	Urinary polyomavirus	Intestinal infection	3	707	Alive	Intestine (2, +226d)
10	Skin and liver	IV	NR	–	Intestinal infection Fungal pneumonia	4	173	Dead	–
11	Liver	IV	NR	CMV	Sepsis	4	53	Dead	–
12	None	III	NR	CMV	Pneumonia	4	28	Dead	–
13	Liver	III	CR	CMV	–	4	713	Alive	Liver (1, +42d)
14	None	III	CR	CMV	–	–	31	Dead	–
15	None	III	CR	CMV retinitis	–	4	821	Alive	–
16	Skin	III	CR	–	–	3	527	Alive	–

Table 1 (continued)

#	Additional organ involvement to intestine manifestations	aGVHD grade	Response to combined treatment	Virus reactivation after combined treatment	Infection after combined treatment	Severe cytopenia	Last follow-up (days after response)	Status	GVHD relapse (severity, days after response)
17	Skin and liver	IV	CR	–	–	4	693	Alive	Skin, liver (2,+144d)
18	None	III	CR	CMV	–	3	485	Alive	–
19	Skin and liver	IV	PR	–	Pneumonia	4	524	Alive	–
20	Skin and liver	IV	CR	CMV + EBV	–	4	458	Alive	–
21	None	III	CR	CMV	Intestinal infection	4	384	Alive	–

HSCT hematopoietic stem cell transplantation, aGVHD acute graft-versus-host disease, AML acute myelogenous leukemia, MDS myelodysplastic syndrome, ALL acute lymphoblastic leukemia, SAA severe aplastic anemia, HAL hybrid acute leukemia, CR complete response, NR no response, PD progressive disease, MUD matched unrelated donor, HRD haplo-identical related donor, BM bone marrow, PB peripheral blood, CSA cyclosporine A, MMF mycophenolate mofetil, MTX methotrexate, FK506 tacrolimus, ATG anti-thymocyte globulin

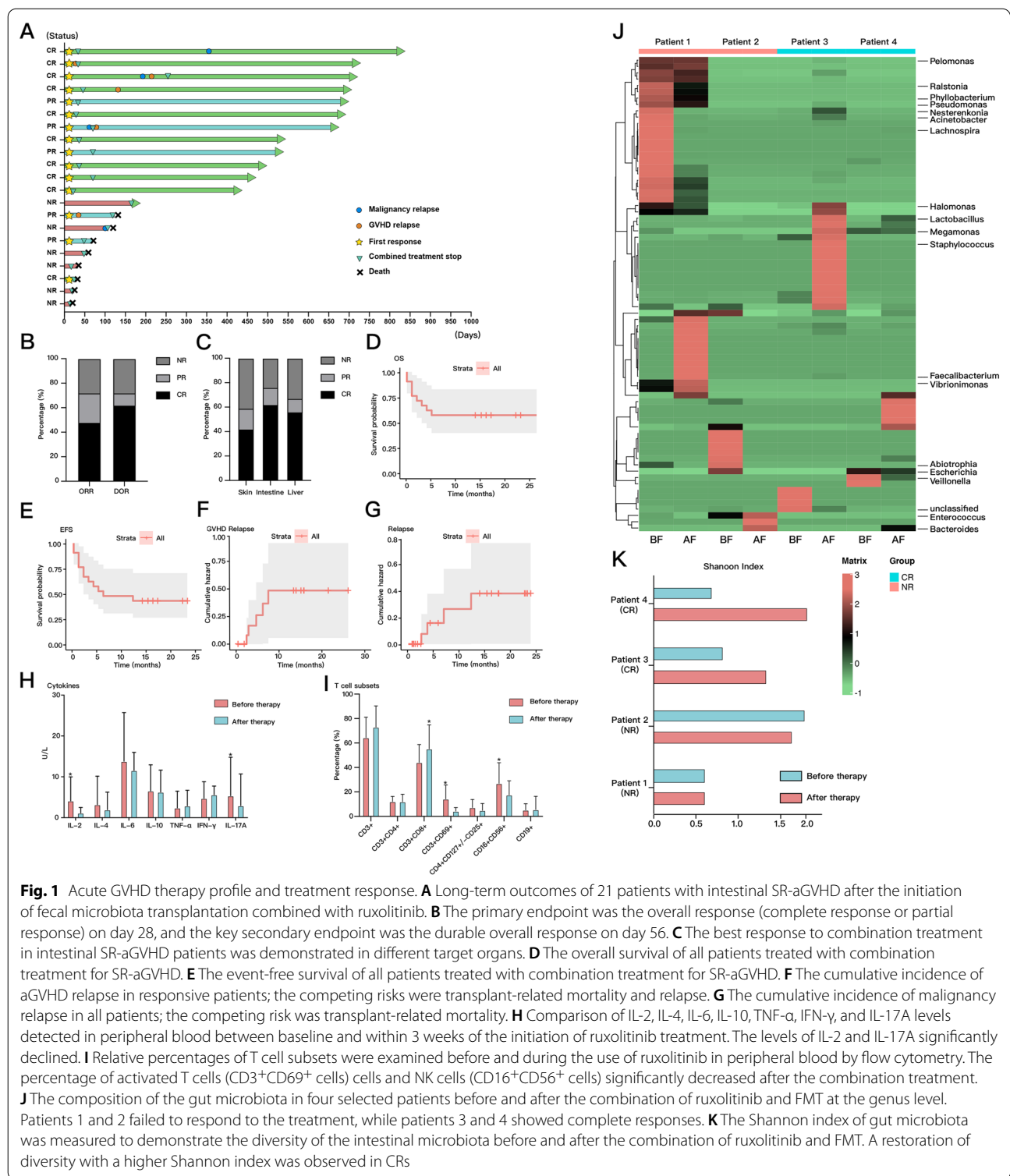
III–IV intestinal SR-aGVHD after HSCT between November 2017 and July 2019. The inclusion criteria included: (1) age from 12 to 60; (2) stable state of disease; (3) grade III–IV intestinal aGVHD; 4) steroid-refractory GVHD defined as the following: disease progression based on organ assessment after at least 3 days of high-dose methylprednisolone (MP) at 2 mg/kg/d, a lack of response without change in visceral GVHD (absence of partial response or better) after 5 days, or exacerbation of acute GVHD or treatment failure during MP taper. Follow-up was continued through July 2019 before the analysis started. The exclusion criteria were uncontrolled infections; inability to swallow tablets and severe organ damage due to reasons other than GVHD. Once diagnosed with steroid-refractory GVHD, patients were given the combined treatment of FMT 2 capsules three times daily for 2–3 days, including 40–50 mg microbiota per full therapy, and oral ruxolitinib with an initial dose of 5 mg twice daily. The median delay between initiation of ruxolitinib and first FMT was 4 days, while the max was 11 days. Frozen fecal microbiota was obtained from China fmtBank (Nanjing, China) and transferred into capsules for swallowing. Our study obtained ethics approval from the Ethics Review Committee of our institution and was conducted in accordance with the Declaration of Helsinki. All patients were provided with written informed consent for participation.

GVHD grading was assessed routinely using standardized criteria per MAGIC guidelines [7]. Peripheral blood counts, infections, and virus reactivations were also monitored closely. Treatment responses were defined as complete response (CR), partial response (PR), or treatment failure (NR). A CR was defined as the absence of any symptoms related to GVHD. A PR was defined as

the improvement of at least one grade in the severity of aGVHD in at least one site and without deterioration in any other organ. Treatment failure was defined as the absence of improvement, deterioration in any organ, or the development of new GVHD symptoms. Overall response rate (ORR), durable overall response (DOR), time to first response, overall survival (OS), event-free survival (EFS), malignancy relapse rate, GVHD relapse rate, and treatment-related adverse events were assessed. Levels of cytokines and the percentages of lymphocytes were measured before treatment and at the best response time.

A cohort of 21 patients, average of 29 years old (range: 15–59), received FMT plus ruxolitinib as a further treatment for grade III–IV intestinal SR-aGVHD in our center between November 2017 and July 2019 (Table 1). Most enrolled patients manifested GVHD in at least 2 organs, including skin rash (11/21, 52.4%) and elevated bilirubin (10/21, 47.6%), despite enteritis with diarrhea. Their characteristics and therapy-related profiles are shown in Table 1 and Fig. 1A.

The ORR on day 28 was 71.4% (95% CI 50.4–92.5%), including 10 CRs and 5 PRs, with a median time of 10 days to achieve the first response. The DOR at day 56 in responders was 80% (Fig. 1B). The median duration of follow-up was 15.7 months. The median duration of steroid tapering to half dose was 14 days. A higher overall response rate (76.2%) was observed in patients with intestinal involvement among distinct target organs (Fig. 1C). The estimated 6-month OS was 57.1% (95% CI: 35.9–78.3%), while EFS was 52.4% (95% CI: 21.7%–64.1%) (Fig. 1D, E). GVHD relapse rate was 33.3% in responders, among whom three patients experienced chronic GVHD (Fig. 1F). Meanwhile, malignancy relapse was observed in four patients at the last



follow-up (Fig. 1G). Viral reactivations (61.9%), bacterial infections (28.6%), and severe cytopenia (grades 3–4, 81%) were the most frequent adverse events observed in our study.

In inflammatory cytokines analysis, we observed significant declines in IL-2 and IL-17A and similar trends in IL-4, IL-6, and IL-10 following the combined treatment compared to the baseline values (Fig. 1H). Additionally,

the percentage of activated T cells and NK cells decreased at the same time (Fig. 1I). We collected data on the temporal microbiota dynamics of four patients (two CRs and NRs respectively). The percentage of beneficial bacteria, such as *Lactobacillus*, increased in CRs; whereas *Escherichia*, which was reported to be strongly correlated with GVHD in a mouse model, reduced after the treatment (Fig. 1J) [1, 8, 9]. The diversity of the intestinal microbiota was improved in responders, with an apparent increase in the Shannon index (Fig. 1K). Furthermore, the levels of inflammatory cytokines and the percentages of activated T cells declined, while regulatory T cells increased in answer to the combined treatment when compared to the baseline levels in patient 3 with a CR (Additional file 1: Figure S1 C, D).

Our study further underlined the additive effect of FMT with ruxolitinib in the salvage treatment of intestinal SR-aGVHD, supported by a high ORR of 71.4% and impressive outcomes. Randomized controlled trials are needed to be conducted to demonstrate the efficiency and safety of the combined treatment. We hope the modifications of protocols for combined treatment with FMT and ruxolitinib could be taken into consideration.

Abbreviations

aGVHD: Acute graft-versus-host disease; HSCT: Hematopoietic stem cell transplantation; SR-aGVHD: Steroid-refractory aGVHD; FMT: Fecal microbiota transplantation; MP: Methylprednisolone; ORR: Overall response rate; DOR: Durable overall response; OS: Overall survival; EFS: Event-free survival; CR: Complete response; PR: Partial response.

Supplementary Information

The online version contains supplementary material available at <https://doi.org/10.1186/s40164-022-00350-6>.

Additional file 1: Figure S1. Comparison of inflammatory cytokines and T cell subsets in the four selected patients. **A** Comparison of IL-2, IL-4, IL-6, IL-10, TNF- α , IFN- γ , and IL-17A levels detected in peripheral blood between baseline and the following 4 weeks after the initiation of combined treatment in Patient 1 with no response. The level of cytokines showed no significant change at 3 weeks and extraordinarily increased at the last follow-up, ending with death. **B** Relative percentages of T cell subsets were examined before and 4 weeks after the initiation of combination therapy in peripheral blood by flow cytometry in Patient 1 with no response. The percentage of CD3+CD69+ cells showed no difference, while the percentage of CD4+CD127+/-CD25+ cells declined to zero. **C** Comparison of IL-2, IL-4, IL-6, IL-10, TNF- α , IFN- γ , and IL-17A levels detected in peripheral blood between baseline and the following 4 weeks after the initiation of combined treatment in Patient 3 with a complete response. The level of cytokines significantly declined. **D** Relative percentages of T cell subsets were examined before and 4 weeks after the initiation of combination therapy in peripheral blood by flow cytometry in Patient 3 with a complete response. The percentage of CD3+CD69+ cells declined coincidentally, while the percentage of CD4+CD127+/-CD25+ cells increased to a remarkably high level.

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Author contributions

Author statement: DW performed the research. YX designed the research study. XM and XQ contributed to the collection of data. JQ analyzed the data. JQ and YZ. discussed and interpreted the results. YL wrote the manuscript. All the authors have read and approved the final manuscript.

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Availability of data and materials

The datasets used and analyzed during the current study are available from the corresponding author upon reasonable request.

Declarations

Ethics approval and consent to participate

This study was conducted in accordance with the Declaration of Helsinki and was approved by the Ethics Review Committee of the First Affiliated Hospital of Soochow University. All patients were provided with written informed consent for participation.

Competing interests

The authors declare no conflicts of interest.

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References

- Kakahana K, Fujioka Y, Suda W, et al. Fecal microbiota transplantation for patients with steroid-resistant acute graft-versus-host disease of the gut. *Blood*. 2016;128(16):2083–8. <https://doi.org/10.1182/blood-2016-05-717652>.
- Bilinski J, Jasinski M, Tomaszewska A, et al. Fecal microbiota transplantation with ruxolitinib as a treatment modality for steroid-refractory/dependent acute, gastrointestinal graft-versus-host disease: a case series. *Am J Hematol*. 2021;96(12):E461–3. <https://doi.org/10.1002/ajh.26365>.
- <https://clinicaltrials.gov/ct2/show/NCT04269850>
- <https://clinicaltrials.gov/ct2/show/NCT03148743>
- Qi X, Li X, Zhao Y, et al. Treating steroid refractory intestinal acute graft-versus-host disease with fecal microbiota transplantation: a pilot study. *Front Immunol*. 2018;9:2195. <https://doi.org/10.3389/fimmu.2018.02195>.
- Zhao Y, Li X, Zhou Y, et al. Safety and efficacy of fecal microbiota transplantation for grade IV steroid refractory GI-GvHD patients: interim results from FMT2017002 Trial. *Front Immunol*. 2021;12: 678476. <https://doi.org/10.3389/fimmu.2021.678476>.
- Schoemans HM, Lee SJ, Ferrara JL, et al. EBMT-NIH-CIBMTR task force position statement on standardized terminology & guidance for graft-versus-host disease assessment. *Bone Marrow Transplant*. 2018;53(11):1401–15. <https://doi.org/10.1038/s41409-018-0204-7>.

8. Jenq RR, Ubeda C, Taur Y, et al. Regulation of intestinal inflammation by microbiota following allogeneic bone marrow transplantation. *J Exp Med*. 2012;209(5):903–11. <https://doi.org/10.1084/jem.20112408>.
9. Kaysen A, Heintz-Buschart A, Muller EEL, et al. Integrated meta-omic analyses of the gastrointestinal tract microbiome in patients undergoing allogeneic hematopoietic stem cell transplantation. *Transl Res*. 2017;186:79–94 e1. <https://doi.org/10.1016/j.trsl.2017.06.008>.

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